Nutritional Deficiency & Impact on Health

Chapter 5

Micronutrients Deficiencies in Early Life and Impact on Long-term Health

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Nutrition in early life is of extreme important due to direct correlation with long-term increased risk of non-communicable diseases (NCDs). The fetal and postnatal level of micronutrients and polyunsaturated fatty acids (PUFAs) may alter metabolism, organ growth, development and function leading to increased risk of cardio metabolic diseases, obesity, renal disorders, respiratory disorders, metabolic syndrome, and, eventually to type 2 diabetes and cardiovascular diseases. This chapter is focused on the complexed effects of micronutrients (vitamins and minerals) and PUFAs deficiencies on the health and discuss the importance of supplementation and education programs as a health policy strategy.

Keywords: Micronutrients deficiencies; PUFAs (polyunsaturated fatty acids) deficiencies; Early life nutrition; Long-term health; Nutrition literacy.

1. Introduction

The intake of suitable macronutrient and the impact on growth and development as well on wellness is well-known. It is sustained by a balanced and diversified diet. Nevertheless, the adequate fetal and postnatal intake of vitamins could impact significantly early life and longterm effects on predisposition to several health outcomes (**Figure 1**).

VITAMINS	COMMON FOOD SOURCES	MEDICAL IMPLICATIONS OF DEFICIENCY				
Water-soluble vitamins						
Vitamin B ₁ (thiamine)	Enriched cereals and breads; unrefined grain; pork; legumes, seeds nuts	Beri-beri : Edema, anorexia, weight loss; apathy, decrease in short-term memory, confusion; irritability; muscle weakness; an enlarged heart				
Vitamin B ₂ (riboflavin)	Dairy products, fortified cereals, meats, poultry, fish, legumes	Ariboflavinosis: Sore throat, hyperemia, edema of oral mucosal membranes, cheilosis, angular stomatitis, glossitis, magenta tongue, seborrheic dermatitis, normochromic normocytic anemia				
Vitamin B ₃ (niacin)	Meat: chicken, beef, fish; enriched cereals or whole grains; most foods	Pellagra : Pigmented rush in areas exposed to sunlight; vomiting; constipation or diarrhea; bright red tongue; neurologic symptoms.				
Vitamin B5 (pantothenic acid)	Wide distributions in foods, especially animal tissues; whole grain cereals; legumes	Irritability and restlessness, fatigue, apathy, malaise, gastrointestinal symptoms, neurological symptoms				
Vitamin B6 (pyridoxine)	Chicken, fish, pork, eggs, fortified cereals, un-milled rice, oats, starchy vegetables, non-citrus fruits; peanuts, walnuts	Seborrheic dermatitis, microcytic anemia, epileptiform convulsions, depression and confusion				
Vitamin B7 (biotin)	Liver Egg yolk	Conjunctivitis; central nervous system abnormalities, glossitis, alopecia, dry, scaly dermatitis				
Vitamin B9 (folic acid)	Citrus fruits, dark green vegetables, fortified cereals and breads, legumes	Impaired cell division and grown; megaloblastic anemia , neural tubes defects				
Vitamin B ₁₂ (cobalamin)	Animal products, fortified cereals	Megaloblastic anemia, neurologic symptoms				
Vitamin C	Citrus fruits, potatoes, peppers, broccoli, spinach, strawberries	Scurvy, defective collagen formation leading to subcutaneous hemorrhage: aching bones, joints, and muscle in adults, rigid position and pain in infants				
Fat-soluble vitamins						
Vitamin A	Carrots; dark green and leafy vegetables, sweet potatoes and squash, broccoli	Night blindness; xerophthalmia ; keratinisation of epithelium in gastro-intestinal system; respiratory and genitourinary tract, skin becomes dry and scaly				
Vitamin D	Fortified milk; exposure of skin to sunlight	Rickets in children; inadequate bone mineralization (osteomalacia)				
Vitamin E	Vegetable oils, margarine; wheat germ; nuts, green leafy vegetables	Muscular dystrophy, neurologic abnormalities				
Vitamin K	Green leafy vegetables; cabbage family bacterial flora of intestine	Defective blood coagulation, hemorrhagic anemia of newborn				

Figure 1: Synthesis of vitamins, common food sources and medical implications [1-7].

Whole blood collected on an anticoagulant substance can be separated by centrifugation into blood plasma and cells.

About 10% of blood plasma are solutes, of which: a) about 70% are plasma proteins; b) 30% are: small organic molecules, inorganic salts, and many other substances that even in small amounts are essentials: enzymes, hormones, other metabolites, and important nutrients as vitamins and trace elements (**Figure 2**).

Measurements of the concentrations of components in blood plasma are very important in the diagnosis and treatment of many diseases, and should be a current diagnosis paraclinical test that help on patient management.



Figure 2: The composition of blood; vitamins and trace elements in a quantitative representation (modified after Lehninger Principles of Biochemistry).

2. Deficiencies of Vitamins Fetal and Neonatal

2.1. Deficiencies of vitamin B complex (Oana Lelia Pop)

B vitamins play a critical role in maintaining a proper function of the human body that is transposed into good health and well-being. Directly affected by the levels of B vitamins are processes as: energy delivery, intellectual functions, and cell metabolism. The presence of the vitamin B complex in fetal and postnatal life gives it an important role, especially in the nourishment of the premature neonates [8]. **Figure 3** shows the vitamin B complex and some natural sources, with their content in 100 g edible portion of food. The fetus feeds from the mother through the umbilical cord after being filtered through the placenta, so the mother's diet and her stocks in B vitamins are essential nutrients [9]. Furthermore, health authorities, including the World Health Organization (WHO), endorse breastfeeding exclusively for six months [10]. The role of vitamin B complex in the normal growth of the fetus and the newborn baby is more or less defined in the scientific papers. The growth and evolution of babies are influenced by the presence of the correct dosage of B vitamins in their diet [11].

Thiamin, or B1, is acting as a coenzyme in the metabolism of carbohydrates and branched-chain amino acids. This vitamin is important in maintenance of nerve membrane

function, and the synthesis of myelin and several types of neurotransmitters (acetylcholine, serotonin, and amino acids). Deficiencies in B1 vitamin is linked to beriberi disease. This disease was registered in infants at 3 to 4 weeks of age from mothers with beriberi. Table 1 offers the recommended adequate intake (AI) for babies breastfed (up to 6 months), babies from 7 to 12 months, and for the pregnant woman.



Figure 3: Vitamin B complex and food sources per 100 g.

Thus, only a well-nurtured mother will be able to ensure the B1 vitamin amount needed for her infant [12]. No significant differences were observed in the milk of lactating women with thiamine supplements and lactating well-nourished mothers [13].

The determination of B2 vitamin or riboflavin AI was established having in mind the correlation between this vitamin amount in human milk and the milk daily intake. Because most plant and animal-based foods contain at least small amounts of riboflavin [14] covering the AI, the estimated average requirements (EAR) is not a great issue for the lactating mothers and food feed babies (7-12 months). Folate (B9), niacin (B3), riboflavin (B2), pyridoxine (B6) and B12 (cobalamin) correlated with zinc ingestion are involved in carbon metabolism. Based on these, B complex are important in early gestation for early cell proliferation, growth, and protein synthesis [15]. Signs of different deficiencies in infants can be seen during pregnancy. Fetal resorption was observed in rats and mice with riboflavin deficiency, without a relevant correlation to humans. Recurrent cleft lip and cleft palate in siblings were associated with riboflavin, vitamin A and folic acid (B9) deficiencies [16].

B5 vitamin is an essential nutrient for humans, necessary for the biosynthesis of coenzyme A, which is needed in a vast range of biological processes, such as fatty acids metabolism [17].

Correlations between the newborn initial weight loss, the rate of return to the birth weight, and the weight at discharge are done in correlation with pyridoxine or vitamin B6. The most common clinical symptoms of B6 deficiency are seborrheic dermatitis, microcytic ane-

mia, epileptiform convulsions, and depression, facts that can affect the mother and the infant as well [13].

Mammals obtain B7 or biotin from food products (registering low amounts in fruits and high amounts in animal organs) or from gut bacteria. Deficiency in biotin for infants and pregnant women are rarely reported. Results showed that diet supplementation with10 mg/day of biotin during the ninth month of pregnancy induce no adverse effects to the mother or infant [11].

The folic acid (B9 vitamin) deficiency is correlated with the coexisting iron or vitamin B12 in an adequate amount. During pregnancy, folate requirements are higher due to the intense cell division and metabolism related to placental and fetal evolution, uterine expansion, and maternal blood volume enlargement. Women of delivering age, i.e., those becoming pregnant, and pregnant women in the first trimester are recommended to increase the folic acid intake per day from supplements. This recommendation is associated with effective action in neural tube defects risk reduction [18,19].

Vitamin B12 or cobalamin, together with folate, are mandatory cofactors in the synthesis of RNA and DNA. Cobalamin is needed for maintaining the nervous system functionality. Therefore, cobalamin is critical to the development during the early years of life [20,21]. Cobalamin stock in utero is an important determinant of cobalamin status in the newborn and during infancy, and there is an effective connection between maternal and newborn cobalamin status. Still, little data exist regarding the relationship between maternal factors influence infant vitamin status after the newborn period or how newborn folate and cobalamin status is related to the situation later in infancy and childhood [22].

Deviteration	Pregnant women		0-6 months infant	7-12 months	
Bvitamin	EAR*	RDA**	AI***	infant AI***	
B_1 thiamine	1.2 mg/day	1.4 mg/day	0.2 mg/day	0.3 mg/day	
B ₂ riboflavin	1.2 mg/day	1.4 mg/day	0.3 mg/day	0.4 mg/day	
B ₃ niacin	14 mg/day	18 mg/day	2 mg/day	4 mg/day	
B ₅ pantothenic acid	6 mg/day***	-	1.7 mg/day	1.8 mg/day	
B ₆ pyridoxine	1.6 mg/day	1.9 mg/day	0.1 mg/day	0.3 mg/day	
B ₇ biotin	30 µg/day***	-	5 μg/day	6 μg/day	
B ₉ folate	520 µg/day	600 μg/day	65 μg/day	80 µg/day	
B ₁₂ cobalamin	2.2 µg /day	2.6 μg /day	0.4 μg/day	0.5 μg/day	

Tabel 1: Dietary Reference Intakes for the vitamin B complex for pregnant woman and for infants of 0-6 months and7-12 months [23].

*EAR - estimated average requirements; ***AI- adequate intake

Studies regarding the deficiencies in early life and impact on the long-term health of most of the B vitamins are limited, with fluctuations regarding folate and cobalamin, which are

the most studied. Thus, close awareness should be given to the identification of indicators on which to base B complex supplementation. Nearly all of the B complex vitamin deficiencies are, in most cases, reversible and can be recovered in days up to 2-3 years (i.e. B12). However, because of their role in critical deficiency illnesses such as pellagra (a lack of niacin) and anemia (riboflavin and other B vitamins), as well as skin and mouth lesions, these B vitamin complex are added to breakfast designated cereals, to milk formula and supplementary foods. B vitamin requirements can easily recovered with adequate and equilibrate nutrition when no other genetic or malabsorption illnesses are correlated or special diets adopted (i.e. vegan).

2.2. Deficiencies of Biotin (Romana Vulturar)

Biotin (vitamin H /vitamin B7) is a water-soluble vitamin and has a key role in energy metabolism and regulation of oxidative stress, being a crucial cofactor for five carboxylases involved in gluconeogenesis, fatty acid and amino acid metabolism [24,25]. New evidence shows a vital role of biotin in chromatin structure, gene expression and genome stability [24,26,27].

Mammals obtain biotin from food (egg yolk, liver, wheat, oats, spinach, mushrooms, rice), or from gut bacteria [24, 26,27].

Causes of biotin deficiency (BD):

1. Genetics [2, 3]: few Inborn Errors of Metabolism (IEM) due to enzyme deficiencies: holocarboxylase synthetase or biotinidase, that if untreated give severe BD. Biotinidase screening should be part of the workup of newborns/children showing clinical features, and the clinicians should follow these patients, takes months to reverse the symptoms, but with good compliance, the outcomes are good.

2. Acquired [2, 3, 24, 27, 28]:

a. Severely malnourished children in developing countries and through the intake of modified milk without biotin supplementation;

b. Alteration in microbiota due to broad-spectrum-antibiotics treatment or inflammatory bowel disease,

c. Drug-vitamin interaction: patients receiving antiepileptics (carbamazepine, valproate, phenytoin, phenobarbital) or isotretinoin for acne treatment, or parenteral nutrition; biotin requirements may increase during these therapies.

d. Elderly, smokers, excessive consumption of alcohol, consumption of large amounts of raw egg whites (avidin binds biotin) reduces biotin absorption.

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e. Marginal BD is common in pregnancy (about half of the pregnant women in USA); the negative statistically significant correlation between hyperemesis gravidarum severity and serum biotin levels was noted. Likewise, lactation can lead to an increased demand for biotin.

Regarding the pathophysiology of BD, this can leads to several abnormalities, mainly dermal and neurological dysfunctions. Biotin regulates immunological and inflammatory functions, playing a key role in the function of natural killer lymphocytes, in B- and T-cell immunity. In BD there are increasing levels of IL-1-beta and proinflammatory cytokines TNF-alpha [24,25].

Dermal abnormalities: hair loss (alopecia) and periorificial (eyes, nose, mouth) dermatitis with a scaly, red rash similar to that of zinc deficiency. Patients may also develop conjunctivitis and skin infections [2,29].

Neurological symptoms: hypotonia, seizures, ataxia, mental retardation, numbness of the extremities, and developmental delay in children. The patient may also show depression, lethargy, a history of hallucinations, ketolactic acidosis, organic aciduria. Initial clinical symptoms of acquired BD include gradual hair loss, dry skin, lesions on the feet and legs. Infants may initially show mild scaly dermatitis on the face similar to soaps-dermatitis-rash. In adults (after a few weeks of having a raw-egg-diet) desquamative dermatitis, anorexia, lethargy, hyperaesthesia are observed; in these cases, administration of biotin relieved symptoms in five days. Individuals with hereditary disorders of BD may show impaired immune function with increased susceptibility to candidiasis. Biotinidase deficiency typically shows symptoms between the age of one week to more than one year and may associates hearing loss, optic atrophy [3,26].

For treatment/ management, oral biotin supplements have high bioavailability, and patients usually respond well to large doses of biotin (5 mg/day regardless of the etiology of BD). Early intervention with lifelong biotin doses (5-20 mg/day) can treat/prevent clinical signs of biotinidase deficiency; failure to manage biotinidase deficiency at an early stage can cause irreversible neurodevelopmental delay and autistic behavior [3,26,30].

The differential diagnosis: a) sodium-dependent multivitamin transporter defect, that is another IEM [31], b) acrodermatitis enteropathica (given by disorders in zinc metabolism) [27].

Regarding the fetal development, evidence shows that lack of biotin is teratogenic in animal models (malformations: mainly cleft palate, micrognathia, micromelia) [26,27,32].

2.3. Deficiencies of Folic acid (Adriana Fodor)

Folate is a water-soluble B vitamin that occurs naturally in food. Folic acid is the synthetic form, with increased bioavailability, used for food fortification and in supplements.

Folates function as cofactors within C1 metabolism, required for DNA and RNA biosynthesis, amino acid and lipids metabolism and methylation processes. Thus, folate deficiency will affect more the cells that rapidly divide, including the progenitors of red blood cells, thereby producing megaloblastic anemia (immature, enlarged blood cells), other bone marrow cells, leading to leukopenia and thrombocytopenia or fetal cells, leading to low birth weight, preterm delivery and fetal growth retardation.

During embryogenesis, the neural tube has increased need of folate for cell differentiation, growth, and closure to form the spinal cord and brain. Folate deficiency during the periconceptional period can induce neural tube defects (NTDs), which are the most frequent human malformations occurring during pregnancy. Conclusive evidence from randomized controlled trials has shown that folic acid supplementation in the periconceptional period unequivocally reduces the occurrence of NTDs, and significantly reduces the risk for other congenital malformations like heart defects and orofacial clefts. For the prevention of NTDs, women are recommended to take 400 μ g/d folic acid as a supplement from preconception until the end of the first trimester of pregnancy [33]. However, the evidence suggests that the current recommendations are largely ineffective because of the poor compliance of women with folic acid supplementation as recommended before and in early pregnancy [34]. In contrast, when mandatory folic acid fortification is undertaken widely on population, it results in marked reductions in NTDs and very low rates of anemia secondary to folate deficiency [35]. Over eighty countries worldwide to date (including the USA, Canada and Australia) have regulations for the mandatory fortification of staple foods with folic acid in order to prevent NTDs.

As folate is required for the re-methylation of homocysteine to methionine, a typical consequence of folate deficiency is an elevation in plasma homocysteine, which, in turn, is implicated in the etiology of cardiovascular diseases [36].

Emerging evidence from candidate gene approach studies links maternal folate during pregnancy with DNA methylation in offspring genes involved in neurodevelopment and cognitive function in childhood [37]. One randomized trial to date has shown that folic acid supplementation in pregnancy led to significant changes in DNA methylation in genes related to brain development, IGF2 and BDNF [38].

Apart from maternal folate status during pregnancy, the child's folate status also seems to impact health. A prospective cohort study in 2922 children has shown that high dietary intake of folic acid at the age of 1 year was associated with a lower body weight at the age of 6 years [39].

2.4. Deficiencies of vitamin C (Angela Cozma)

Vitamin C, also known as ascorbic acid (AsA) is recognized as a vital dietary micronutrient based on its ability to prevent scurvy in humans. While this condition is rare in the Western world, AsA deficiency, defined by a plasma concentration $< 23 \mu mol/l$, is surprisingly common [40].

AsA plays a major role in defense against increased oxidative stress during pregnancy [41]. AsA levels decrease during pregnancy, due to physiological changes in pregnancy and inadequate intake [42]. Pregnancy is associated with increased susceptibility to oxidative stress. The antioxidant deficiencies during pregnancy and placental oxidant-antioxidant imbalance may affect the development of the fetoplacental unit or the eventual offspring [43]. There are many studies regarding AsA deficiency during pregnancy and low birth weight and preterm delivery [44-46].

The increased consumption of fruit and vegetables, an excellent source of AsA, during pregnancy has been shown to be positively associated with birth weight [47,48].

Consistent findings from experimental animal models of both AsA depletion and deficiency have suggested that AsA -is playing a crucial role in the brain, particularly during development [49-51].

In humans, studies have shown that poor maternal AsA status results in increased fetal oxidative stress, impaired implantation and increased risk of complications including preeclampsia [52-54]. Women with pre-eclampsia have been shown to have reduced levels of AsA and several studies have investigated a potential effect of AsA supplementation [55]. It is not clear to what extent AsA supplementation may ameliorate this risk.

Recently, Takeshita examined the effect of AsA deficiency on the concentration of tetrahydrobiopterin (BH₄) using ODS rats, which are defective in the gene for AsA synthesis. AsA deficiency determines a decrease in the monoamine levels in the brain and plays an important role in the pathophysiology of neuropsychiatric and cardiovascular disorders through alteration in BH₄ metabolism [56].

On the other hand, it has been shown that AsA modulates ten-eleven translocation (TET) activity, an enzyme with a role in DNA demethylation. DiTroia has shown that maternal AsA is required for proper DNA demethylation, and the development of female fetal germ cells in a mouse model with AsA deficiency leads to an aberrant DNA methylation. Maternal AsA deficiency does not affect overall embryonic development but leads to reduced numbers of germ cells, delayed meiosis and reduced fecundity in adult offspring. The author concludes that deficiency in AsA during gestation partially recapitulates loss of TET1, and provide a

potential intergenerational mechanism for adjusting fecundity to environmental conditions [57].

2.5. Deficiencies of vitamin A (Adina Chiş)

Vitamin A (all-trans-retinol) deficiency (VAD) is a major health issue worldwide, especially in low/middle-income countries; risk groups for VAD have been identified even in developed countries [58]. The VAD may have serious consequences at any age; when it appears during pregnancy and early childhood, the consequences can be severe for maternal health, but mainly for the fetus.

Low intake of vitamin A, especially during the third quarter of pregnancy, can lead to fetal skeleton malformations, impairments in the ocular and immune fetal systems development; in more severe cases of maternal deprivation, premature birth or even fetus death can occur [59,60].

Without fully understanding the pathophysiology, studies have shown that in neonate fed with low-vitamin A milk, the immune system is affected: an increased risk for respiratory tract infections and complications during viral infections (measles) can occur.

During the childhood, the VAD is associated with xerophthalmia that, untreated, can progress to nyctalopia; later in life, if this deficiency is untreated will affect: cell differentiation, growth and reproduction, will increase the risk of infection and, even the mortality [58].

Furthermore, animal studies support the involvement of VAD in the pathogenesis of certain diseases; depending on the severity of VAD, were observed anomalies in the development of eyes, brain, kidneys, lungs, and may cause even death and fetus resorption. There are also studies that have shown that the VAD during the pregnancy was associated with the development of diabetes later in life [4, 58]

2.6. Deficiencies of vitamin D3 (Adela Viviana Sitar Tãut)

Vitamin D deficiency (VDD) represents a public health problem, being observed worldwide in pregnant women and their newborns [5, 61]. An important proportion of pregnant women (prevalence ranged 18-84%) [5, 6, 62, 63], respectively of newborns [6] presents VDD. According to the most experts, vitamin D deficiency is defined as a 25-hydroxyvitamin D level less than 20 ng/ml [5, 64], but increased values are recommended by Endocrine Society during pregnancy (at least 30 mg/ml) [5]. Many studies showed a positive correlation between maternal and fetus/newborn's vitamin D levels [61], the fetus being dependent on maternal D vitamin level [62,65] (**Table 2**).

Risk factors in developing vitamin D deficiency are extremely various; there are

mentioned causes like receiving insufficient vitamins [63], low intake of fortified food and low compliance of supplementation [63], black race [6], body covering [63], winter birth [6] living in cold climates or northern latitudes [66,67], inadequate sunlight exposure [5], maternal deficiency in relationship or not with diet (vegetarians) [5,6,66], maternal obesity [6, 68] (**Table 2**).

Recent studies have shown that vitamin D deficiency has a major impact not just only on skeletal development and bone mineralization [5,6,62,63,65,69,70], but also being associated with a broader range of adverse infant and maternal health outcomes (on a short and long time) [5,6,71] (**Table 2**).

At maternal level determines: Ref	erences
infertility or implantation failure	[5, 72, 73]
increased risks of primary cesarean delivery	[74, 75]
preterm delivery	[61, 62, 76]
recurrent pregnancy loss (vitamin D inhibits pro-rejection cytokines and fav release of tolerance-promoting cytokines, down-regulate TNF- α , IL-6, and p placental level)	vor the IL-10 at [5, 77, 78, 79]
bacterial vaginosis (insufficient antimicrobial peptide cathelicidin synthesis) [5, 62, 63, 65, 66, 75, 80]
risk of pre-eclampsia (vitamin D influences over RAAS system, over antian factors like FMS-like tyrosine kinase-1 and vascular endothelial growth fac controlling fetal-placental immune responses)	giogenic tor, also [5, 62, 63, 66, 68, 75-77, 81, 82]
risk of gestational diabetes mellitus (vitamin D has positive effects - increas sensitivity and regulate insulin production)	[5, 62, 63, 66, 67, 75-77, 82-86]
risk of postpartum depression, periodontal disease	[75]
Intrauterine determines:	
intrauterine growth restriction	[5, 62, 66, 75-77]
low birth weight, length	[61-63 66, 71, 87]
head circumference	[66]
increased risk of small-for-gestational-age	[7, 62, 71]
development of rickets, skeletal deformities	[7, 63]
increased risk of premature or later in life fracture (abnormality in calcium a phosphate metabolism)	and [62, 75, 88, 89]
pelvic deformity	[5, 7, 75]
alteration of the immune system with an increased risk of lower respiratory infections, early-onset neonatal sepsis (deficiency is associated with dysreg cytokines and immunomodulation)	tract ulation of [62, 63, 69, 75, 90, 91, 92]
decrease induction capacity of antimicrobial peptides - cathelicidin (LL37), beta-3 defensins	beta-2 and [90]
decrease of T helper cells 2 (Th2) differentiation	[90]
altered monocyte and macrophages' response	[92]
diminished activity against bacterial and viral agents	[92]
improper local control of pathogens	[90]

Table 2: Vitamin D Deficiency

impaired fetal and childhood growth and development - slower than normal - of the neonatal cardiovascular system, brain	[5, 70, 76]
lung development	[76, 93, 94]
multiple neonatal respiratory disorders - acute lower respiratory tract infections, transient tachypnea of the newborn, respiratory distress syndrome, bronchopulmonary dysplasia	[5, 62, 94]
vitamin D upregulating some genes involved in lung development – matrix metallopeptidase 9, NF-k light polypeptide gene enhancer in B cells inhibitor, epidermal growth factor receptor, E1A binding protein p300. Also, vitamin D influences alveolar epithelial-mesenchymal interactions	[87, 93]
lipofibroblast proliferation, apoptosis of lung fibroblasts	[94]
increases surfactant synthesis	[87, 91]
the deficit of vitamin D is associated with changes in lung structure and function that can persist into later life	[68, 69, 91, 93, 95]
high neonatal mortality	[61]
susceptibility for later-life diseases - metabolic syndrome, childhood adiposity and obesity, type 1 diabetes or even cardiovascular diseases	[5, 7, 62, 65, 70, 71, 82, 89, 96, 97]
vitamin D deficiency has the potential to program long-term vulnerability to cardiovascular disease (upregulating the renin-angiotensin system, altering normal proliferation and differentiation in the fetal heart, determining hyperplastic cardiomyocyte growth, affecting normal cardiogenesis)	[98, 99]
several case reports of vitamin D deficiency or rickets-associated pediatric cardiomyopathies or congenital heart diseases in offsprings in compromised maternal vitamin D	[99]
possible development of asthma, allergic rhinitis, eczema (in relationship with interaction between vitamin D and developing immune system)	[7, 62, 65, 68, 69, 75, 76, 82, 93, 100, 101, 102]
risk of psychiatric and neurological diseases – VDD can cause abnormal changes in brain morphology and development, possible reductions of neurotrophic factors, nerve growth factors, neuronal differentiation, axonal connectivity	[5, 68, 76]
 involved in: -cellular proliferation, differentiation, neurotransmission and neuroprotection DNA repair mechanisms "serotonin paradox" having anti-inflammatory effects, reducing cytokine associated with cognitive impairment levels 	[67] [67] [67, 103] [67, 104, 105, 106]
deficiency can be responsible for: - seizures or tetany development (due to hypocalcemia) - can play a role in autism or ADHD spectrum disorders - predisposition to schizophrenia, depression or multiple sclerosis	[5, 7, 61, 62, 75, 107] [76, 108, 109, 110, 111] [5, 7, 62, 112, 113]
association between prenatal vitamin D status and global IQ or cognitive development with inconclusive results till now	[76, 114]
risk of retinopathy of prematurity development, being imply the lack of endothelial cells proliferation and angiogenesis' inhibition by vitamin D (via thrombospondin)	
pediatric oncology patients, the deficit of vitamin D being frequently met in oncology patients. Future studies are needed to examine causality, to evaluate the frequency of vitamin D receptor (VDR) polymorphisms in children diagnosed with cancer, to establish the real temporal relationship between vitamin D deficiency and cancer	[116]

2.7. Deficiencies of vitamin E (Olga Orãșan)

The vitamin E (or alpha-tocopherol) is found in a variety of tissues, being lipid-soluble, and taken up by the body in a wide variety of ways. The content of alpha-tocopherol in the body of the fetus remains at 3-7 mg/kg throughout gestation [117].

Several genetic disorders lead to vitamin E deficiency, including:

- Mutations in the gene encoding hepatic alpha-tocopherol transfer protein, with symptoms similar to Friedreich ataxia [118-122].
- Abetalipoproteinemia, due to mutations in the microsomal triglyceride transfer protein associated with very low or absent levels of low-density lipoprotein and fat-soluble vitamin deficiencies. Symptoms include progressive ataxia, sensory-motor neuropathy, and vision impairment with retinitis pigmentosa.

In premature infants, vitamin E deficiency may cause a hemolytic anemia. Congenital hemolytic disorders such as thalassemia, sickle cell anemia, glucose-6-phosphate dehydrogenase deficiency, and spherocytosis may be associated with low vitamin E plasma levels [123-126].

In preterm infants, lack of vitamin E intake or fat malabsorption results in edema, thrombocytosis and hemolytic anemia and could eventually result in spinocerebellar degeneration. A tocopherol/total lipid ratio of greater than 0.8mg/g has been recommended to evaluate vitamin E sufficiency [117].

The amount of vitamin E in colostrum and preterm milk is two to three times higher than in mature milk. Formulas for preterm infants should contain at least one IU of vitamin E/g of linoleic acid, 0.6mg of d-alpha-tocopherol and 0.7IU/100kcal [117].

2.8. Deficiencies of vitamin K (Romana Vulturar)

Vitamin K corresponds to a group of fat-soluble compounds and their implication in health is related to vitamin K-dependent-proteins involved in [127-129]:

• **coagulation** (clotting factors II, VII, IX, X);

• cardiovascular system (decreased levels of vitamin K subtypes give arterial calcification)

• **bone development** (osteocalcin, matrix GLA proteins).

The vitamin K1 is predominantly from leafy greens and vegetables, while the main sources of Vitamin K2 are intestinal flora and fermented foods [128].

Vitamin K deficiency is rare in adults (malabsorption syndromes, or those treated with drugs that interfere with vitamin K) [128, 130]. Regarding the newborns, all have reduced Vitamin K at birth (low levels transferred across the placenta and low levels in breast milk).

Vitamin K Deficiency Bleeding (VKDB) is a potentially devastating consequence of vitamin K deficiency in newborns (**Table 3**).

Based on the <i>causes</i> Re	ferences	
 Hereditary-Combined-Vitamin-K-dependent-Clotting-Factors-Deficiency (VKCFD) is extremely rare - genetic recessive disorders with decreased activity of K-dependent proteins. With vitamin K supplementation: good prognosis. Inadequate uptake or secondary to chronic disorders. Context that is drug-related. 		
Based on the timing of the presentation		
 Early VKDB (within 24 h) - when mothers take vitamin K interfering substances (i.e. anticonvulsants); incidence without vitamin K-supplementation has been reported as 12%. Classic VKDB (within the first week) is as a bleeding disorder; in combination with sepsis-induced-bleeding, has 62% fatality rate. Without Vitamin K supplementation, the current incidence is 0.25-1.7%. Late VKDB (between one to twelve weeks) has the worse prognosis, and occurs in 4.4-72 infants /100, 000 births (increased risk in exclusively breastfed infants). 50% of these cases present intracranial hemorrhage (mortality rate is 20-50%). 	[127, 132, 133, 134]	

Table 3: Vitamin K deficiency in newborns.

Vitamin K prophylaxis in newborns: within an hour of birth to prevent severe bleeding - an intramuscular injection or repeated oral doses for a minimum of 6 weeks [127,133].

3. Deficiencies of Minerals (Doina Miere, Lorena Filip, Simona C. Hegheş and Anamaria Cozma-Petruț).

3.1 Deficiencies of Calcium and Magnesium

Calcium (Ca) is essential for both fetal and postnatal bone development. In addition, it is involved in cell membrane functions, blood coagulation, enzyme and hormone actions, muscle contraction and nerve impulse transmission [135]. Physiological adaptations in maternal Ca homeostasis occur during pregnancy to meet the increased demand, but they appear to be independent of maternal Ca supply in populations with adequate Ca intake [136]. Thus, gestational Ca deficiency is rare in Western societies but has been described in low-income countries that have poor nutrition. Hypocalcemia in pregnancy has been linked with pre-eclampsia and intrauterine growth restriction (IUGR) [137]. Furthermore, an association has been suggested between maternal pregnancy-related hypertension and elevated blood pressure among offspring during childhood, adolescence and adulthood [138].

Magnesium (Mg) acts as a co-factor for numerous enzymes and plays an important role in various metabolic processes, neuronal and muscular excitability and vasomotor tone modulation, respectively. Hypomagnesemia during gestation has been associated with hypertensive conditions, preterm delivery and low birth weight [139]. Moreover, intrauterine Mg deficiency has been postulated to alter organ structure and predispose the offspring to metabolic syndrome in later life [140].

The risk of Ca and Mg deficiencies is low in term breast-fed infants in the first six months of life [141]. In contrast, in 6 to 23-month-old children, inadequate complementary feeding has been associated with severe Ca and Mg deficiencies in developing countries. Ca deficiency affects bone mineralization and, consequently, linear growth. Mg deficiency results in anorexia and may indirectly contribute to growth retardation. Linear growth failure not only results in morbidity and mortality but also increases the risk of dyslipidemia, hypertension, and glucose intolerance in later life [142].

3.2. Deficiencies of Zinc

Zinc (Zn) is an essential mineral that plays a key role in cell growth, development, and differentiation having many biological functions including protein synthesis, cellular division and nucleic acid metabolism. Periods of rapid growth such as late pregnancy, infancy and puberty are more vulnerable to zinc deficiency. There are no specialized zinc storage systems in the body. Therefore, a compromised status can develop rapidly and is more common when the main staple foods are high in phytates, or people are following a vegetarian diet or are poorly nourished. Severe zinc deficiency is considered to be rare, while mild or moderate zinc deficiency seems to be more common [143].

It is estimated that 82% of pregnant women worldwide have a zinc intake lower than the recommended dietary intake, and the values may be even higher in developing countries [143]. Even if several earlier reports have shown that maternal zinc deficiency during pregnancy in humans is linked with various adverse pregnant outcomes, recent systematic reviews [144-146] demonstrate that zinc supplementation during pregnancy may reduce risk of preterm birth in a cohort of women who had a previous preterm delivery/ risk of preterm birth. Adequate maternal zinc intake is also critical for optimal brain development in infants and may have long-term consequences, the deficit impairing the process of learning and memory [147,148].

In the early neonatal period, adequate sources of zinc can be obtained from breast milk, but young children's poor nourishment is particularly affected. Zinc deficiency can affect microbiota composition [149] and impair immune function [150] contributing to the global burden of infectious diseases including diarrhea [145], pneumonia and malaria [151,152].

3.3. Deficiencies of Selenium

Selenium (Se) functions as an antioxidant and may play a crucial role in protecting the

fetus from the increased oxidative stress associated with pregnancy. A correlation between Se deficiency during pregnancy and the occurrence of obstetric and perinatal complications has been reported for recurrent miscarriage, preeclampsia, IUGR, preterm delivery and small-for-gestational-age birth weight [153]. Despite some evidence for positive effects of Se supplementation in pregnant women, further studies are required to confirm the benefits and safety of increased Se intake in this population group [154].

Moreover, Se may protect the developing fetal brain from oxidative injury. Se is also involved in thyroid metabolism, with importance for long-term neurodevelopment [155]. A low prenatal Se status has been negatively associated with infant psychomotor score at 6 months of life [156]. In contrast, a positive relationship has been observed between an adequate maternal Se status in pregnancy and cognitive and psychomotor abilities during toddlerhood and middle childhood [157].

Serum Se levels decrease in the first months of life, then show a progressive increase to reach constant values in children over one year of age [158]. Infants are born with Se reserves, but depend as they grow on the Se supplied by breast milk. Se levels in breast milk are high in colostrum, then decrease over lactation and are also strongly influenced by maternal Se status and diet. Furthermore, dietary intake of Se depends on the geographical region and the content of Se in the soil [159]. Nevertheless, overt Se deficiency is relatively rare in term infants. Keshan cardiomyopathy and Kashin-Beck osteoarthropathy are Se-deficient conditions of childhood identified only in certain areas where Se levels in the soil are extremely low [160].

3.4. Deficiencies of Iron

Iron (Fe) plays a key role in hemoglobin and myoglobin synthesis, oxygen transport, and the functioning of iron-dependent enzymes. Iron deficiency (ID) develops when the amount of stored iron is inadequate to meet body requirements and culminates in iron deficiency anemia (IDA) when stored iron is depleted and red cell production is impaired. The risk of ID or IDA is particularly high in pregnancy and infancy [161]. During gestation, iron needs increase compared to the pregestational period to cover maternal red blood cell expansion and the feto-placental growth [162]. After birth, the requirements for iron of the exclusively breastfed term infants are met primarily through the utilization of iron stores because the iron concentration in breast milk is low. At around 6 month of age, iron stores become depleted and introduction of iron-rich foods and even iron supplements is recommended for the rapid growth associated to infancy and toddlerhood [163].

IDA during pregnancy has been associated with preterm birth, small for gestational age, and low birth weight [164]. Although iron supplementation in pregnancy is still controversial in term of safety, an adapted supplemention to the maternal iron status may be recommended [165].

Furthermore, ID during pregnancy, infancy and early childhood may alter neurodevelopment processes such as myelination and neurotransmitter synthesis, resulting in cognitive and behavioral deficits that can persist throughout life, even after iron repletion [166]. Several studies reported long-term cognitive, motor, social-emotional and neurophysiologic abnormalities in both children and adults as a negative effect of ID early in life [167]. In addition, evidence from animal models suggests that maternal ID during pregnancy is associated with offspring obesity and hypertension later in life. However, such evidence is limited in human studies [168].

4. Deficiencies of n3 -, n6 - Polyunsaturated fatty acids (Carmen Ioana Mureșan)

Polyunsaturated fatty acids (PUFAs) can be classified among the omega-3 (n-3) or omega-6 (n-6) families. Some PUFAs cannot be synthesized de novo, and thus are essential fatty acids (EFAs) which need to be taken from the diet during pregnancy, fetal and neonatal period. The parent EFAs are linoleic (LA; 18:2, n-6) and α-linolenic (ALA; 18:3, n-3) acid, which can be further desaturated and elongated (by enzymes) into biologically active longchain PUFAs (LCPUFAs). ALA can be converted into eicosapentaenoic acid (EPA; 20:5, n-3) and docosahexaenoic acid (DHA; 22:6, n-3), and all three represent the most important and common PUFAs found in human milk [169]. DHA has a structural role in the brain grey matter and retinal membranes [170]. LA is metabolized in arachidonic acid (ARA; 20:4, n-6), which can be found in neural tissue [171], and is a precursor of compounds involved in immune response [172]. Still, due to genetic and environmental factors, the fetus and infant can have ineffective enzymes and cannot convert parent EFAs leading to deficiencies [173]. Also, a critical moment which can lead to deficiencies is during the last ten weeks of pregnancy -associated with important brain and nerve tissue development, when fetal demand for EFAs is high [174]. Still, these deficiencies can be surmounted by an adequate maternal diet during late pregnancy and in early neonatal life. Yet, in pathological conditions (i.e., intrauterine growth restriction, diabetes), maternal and fetal levels of LCPUFAs undergo significant changes and deficiencies can also occur [172]. Thus, in these conditions, more attention should be given to the mother's nutritional status. Another issue is the excess of various EFAs, which can diminish the bioavailability of others due to competitive desaturation leading to deficiencies and undesirable consequences such as slower neural transmission times and postnatal growth restriction [175, 176]. The benefits of EFAs supplementation were reported with controversial results. A recent meta-analysis [177] concluded that n-3 PUFAs do not improve visual acuity, growth or language development, whilst motor, cardiovascular health, behavior and immunity were affected.

Also, the Cochrane review [178] showed there is no significant effect of LCPUFAs supplementation. Still, n-3 LCPUFAs consumption could benefit preterm infant development [179-181] and the antiallergic effect in offspring is promising [182-184]. Although more

research is mandatory, the health organizations recognized LCPUFAs as key micronutrients in the first 1000 days of life [185]. The International European Food Safety Authority (EFSA) recommends 100–200 mg/day of DHA in addition to the adequate intake (AI) of 250 mg/day of EPA plus DHA during pregnancy and lactation. The EFSA panel proposes for DHA an AI of 50–100 mg/day for infants and 100 mg for the age of 6–24 months [186].

5. Impact of nutrition literacy of pregnant women and mothers on long-term health (Mãdãlina A. Coman, Bianca O. Duran, Ștefana A. Dobran)

Research in the field of nutrition and health determined that health and nutrition literacy levels of pregnant women directly affect their level of prenatal care and influence the outcomes of pregnancy. Breastfeeding, use of an emergency room, use of medication for infants, and feeding patterns for babies are all strongly influenced by the health and nutrition literacy of the mother [187,188]. Moreover, the nutrition of the mother determines both the physical and mental health of her child and unfavorable events can facilitate vulnerabilities and lead to several cardiovascular and metabolic diseases [189,190]. The body of research on pregnancy, lactation, infancy and early childhood has long established that nutrition has long-lasting effects on later health and disease for both communicable and non-communicable diseases. The phenomenon is referred to as "early metabolic programming of long-term health and disease "[191]. The most common diseases associated with poor nutrition during pregnancy, infancy and early life are cardiovascular diseases and neural tube defects (NTD's), which are characterized by severe defects of the brain and spine, clubfoot, diabetes, obesity and hypertension [192,193]. Nutrition requirements for pregnant women and mothers vary across countries, depending on the guidelines recommended and used. In order for these guidelines to be understood and followed, pregnant women and mothers need to have an adequate level of health literacy and nutrition literacy.

Health literacy is defined as "knowledge, motivation and competencies to access, understand, appraise, and apply health information in order to make judgments and make decisions in everyday life concerning healthcare, disease prevention and health promotion, to maintain or improve quality of life during the life course" [194]. Nutrition literacy is a form of health literacy that encompasses key elements from general health literacy and food literacy constructs [195].

Low levels of health literacy and nutrition literacy are associated with low levels of education and general literacy, and overall poorer health outcomes in individuals, pregnant women are not an exception [196]. Maternal diet during the pregnancy period is considered to be one of the influential factors on child health and development, affecting their long time health. Apart from that, it affects the health condition, both physical and psychological of women going through pregnancy and their early years of motherhood, as demonstrated in the

early chapters of this book. Nutrition education and counseling are strategies that are mostly used around the world in order to improve the health and nutrition literacy of pregnant women and mothers. Counseling about healthy eating and physical activity are extremely important in order to prevent excessive weight gain during pregnancies and in undernourished populations, education on increasing daily energy and protein intake is recommended [197].

World Health Organization (WHO) recommends using a context adapted strategy that is based on the following principles while suggesting further research and development of prenatal programs [198]:

- 1. Education about increasing the diversity and amount of foods consumed.
- 2. Education about adequate weight gain through sufficient and balanced protein and energy intake.
- 3. Education about consistent and continued use of micronutrients supplements, food supplements or fortified foods.

6. Conclusion

The quantity and quality of the micronutrients, vitamins and minerals, or PUFAs from food sources are critical for growth and development as well for biologic activity. The foetal and postnatal deficiencies caused by diet impairments could translate by health effects to infants and potential long-term effects (altering renal function, cardiovascular function, pancreas function, body composition, and pulmonary function). The nutrients needs are challenging for meeting their requirements with diet alone. In response, the recommended use of micronutrients supplements and fortified foods during pregnancy should be a mutual care pregnancy practice. Biotin (vitamin H or B7), vitamin D3, folic acid (vitamin B9), iron, iodine, zinc are extremely important for their potential to further improve infants outcomes beyond micronutrient supplementation. Although, micronutrient supplementation is an effective strategy during pregnancy, the in-depth studies are needed to translate into a public health policy recommendation by WHO, FDA (Food and Drug Administration) and EFSA (European Food and Safety Authority).

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8. References

1. Bird, J.K., et al., Risk of Deficiency in Multiple Concurrent Micronutrients in Children and Adults in the United States. Nutrients, 2017. 9(7):p. 655. https://doi.org/10.3390/nu9070655

2. Ogawa, Y. et al., Biotin Is Required for the Zinc Homeostasis in the Skin. Nutrients, 2019. 11 (4): p.919. https://doi. org/10.3390/nu11040919

3. Saleem F, Soos MP. Biotin Deficiency. [Updated 2020 Jan 20]. In: StatPearls [Internet]. Treasure Island (FL): StatPearls Publishing; 2020 Jan-. Available from: https://www.ncbi.nlm.nih.gov/books/NBK547751/

4. Chien, C.Y., et al., Maternal vitamin A deficiency during pregnancy affects vascularized islet development. Journal of Nutritional Biochemistry, 2016. 36: p. 51–59. https://doi.org/10.1016/j.jnutbio.2016.07.010

5. Heyden, E.L., and S.J. Wimalawansa, Vitamin D: Effects on human reproduction, pregnancy, and fetal well-being. J Steroid Biochem Mol Biol, 2018. 180:p.41–50. https://doi.org/10.1016/j.jsbmb.2017.12.011

6. Merewood, A., et al., Widespread Vitamin D Deficiency in Urban Massachusetts Newborns and Their Mothers. Pediatrics, 2010. 125(4):p.640–647. https://doi.org/10.1542/peds.2009-2158

7. Bianchi, M.L., Vitamin D Homeostasis and Diseases in Pediatrics. Vitamin D in Clinical Medicine, 2018. 50:p.189–201. https://doi.org/10.1159/000486086

8. Yaman, M., and Ö.F. Mızrak, Determination and evaluation of in vitro bioaccessibility of the pyridoxal, pyridoxine, and pyridoxamine forms of vitamin B6 in cereal-based baby foods. Food Chemistry 2019. 298: p.125042. https://doi. org/10.1016/j.foodchem.2019.125042

9. Graulet, B. ; Girard, C.L., Chapter 15 - B Vitamins in Cow Milk: Their Relevance to Human Health, in Dairy in Human Health and Disease Across the Lifespan, R.R. Watson, R.J. Collier, and V.R. Preedy, Editors. 2017, Academic Press. p. 211-224. https://doi.org/10.1016/B978-0-12-809868-4.00015-7

10. World Health Organization. Regional Office for the Western Pacific. (2014). Breastfeeding : a winning goal for life : overcoming obstacles and making an empowered choice. Manila : WHO Regional Office for the Western Pacific. https://apps.who.int/iris/handle/10665/208237

11. Elmadfa, I., and A.L. Meyer, Vitamins for the First 1000 Days: Preparing for Life. International Journal for Vitamin and Nutrition Research 2012, 82, 342-347. https://doi.org/10.1024/0300-9831/a000129

12. Ashworth, A., Efficacy and Effectiveness of Community-Based Treatment of Severe Malnutrition. Food and Nutrition Bulletin, 2006. 27(3_suppl3):p. S24–S48. https://doi.org/10.1177/15648265060273S303

13. Driskell, J.A., Vitamin B-6 requirements of humans. Nutrition Research 1994, 14: p.293-324. https://doi.org/10.1016/S0271-5317(05)80388-4

14. Cataldi, T.R.I.et al., Assessment of riboflavin and flavin content in common food samples by capillary electrophoresis with laser-induced fluorescence detection. Food Chemistry, 2003, 82(2):p. 309-314. https://doi.org/10.1016/S0308-8146(02)00567-8

15. Gernand, A.D., et al., Micronutrient deficiencies in pregnancy worldwide: health effects and prevention. Nat Rev Endocrinol, 2016. 12:p. 274-289. https://doi.org/10.1038/nrendo.2016.37

16. Powers, H.J., Riboflavin (vitamin B-2) and health. The American Journal of Clinical Nutrition, 2003. 77(6): p.1352-1360. https://doi.org/10.1093/ajcn/77.6.1352

17. Martin, F., and E. Campos-Gimenez, Pantothenic Acid (Vitamin B5) in Infant Formula and Adult/ Pediatric Nutritional Formula by Ultra-High Pressure Liquid Chromatography/Tandem Mass Spectrometry Method: Collaborative Study, Final Action 2012.16. Journal of AOAC International, 2015. 98(6):p.1697-1701. https://doi.org/10.5740/jaoacint.15-

18. Lamers, Y., Folate recommendations for pregnancy, lactation, and infancy. Annals of Nutrition and Metabolism, 2011. 59(1):p.32-7. https://doi.org/10.1159/000332073

19. Hermoso, M., et al., Critical micronutrients in pregnancy, lactation, and infancy: considerations on vitamin D, folic acid, and iron, and priorities for future research. Annals of Nutrition and Metabolism, 2011. 59(1):p.5-9. https://doi. org/10.1159/000332062

20. Hay, G., et al., Folate and cobalamin status in relation to breastfeeding and weaning in healthy infants. The American journal of clinical nutrition, 2008. 88(1):p.105-14. https://doi.org/10.1093/ajcn/88.1.105

21. Bjørke-Monsen, A.L., et al., Common metabolic profile in infants indicating impaired cobalamin status responds to cobalamin supplementation. Pediatrics, 2008. 122(1):p.83-91. https://doi.org/10.1542/peds.2007-2716

22. Hay, G., et al., Maternal folate and cobalamin status predicts vitamin status in newborns and 6-month-old infants. The Journal of nutrition, 2010.140(3): p. 557-64. https://doi.org/10.3945/jn.109.117424

23. Institute of Medicine (US) Standing Committee on the Scientific Evaluation of Dietary Reference Intakes. Dietary reference intakes for thiamin, riboflavin, niacin, vitamin B6, folate, vitamin B12, pantothenic acid, biotin, and choline. National Academies Press (US); 1998. ISSN: ISBN-10: 0-309-06411-2

24. Agrawal, S., A. Agrawal, and H.M. Said, Biotin deficiency enhances the inflammatory response of human dendritic cells. American Journal of Physiology-Cell Physiology, 2016. 311(3):p.C386-91. https://doi.org/10.1152/ajpcell.00141.2016

25. Kuroishi, T., Regulation of immunological and inflammatory functions by biotin. Canadian journal of physiology and pharmacology, 2015. 93(12):p.1091-6. https://doi.org/10.1139/cjpp-2014-0460

26. Zempleni, J., Y.I. Hassan, and S.S. Wijeratne, Biotin and biotinidase deficiency. Expert review of endocrinology & metabolism, 2008. 3(6):p.715-24. https://doi.org/10.1586/17446651.3.6.715

27. Maxson & Mitchell. 2016. "Biotin HHS Public Access." Physiology & Behavior 176 (1): 139–48. https://doi. org/10.1016/j.physbeh.2017.03.040.

28. Onder, A.B., et al., Biotin Deficiency in Hyperemesis Gravidarum. Journal of Obstetrics and Gynaecology, 2019. 39
(8): p.1160–63. https://doi.org/10.1080/01443615.2019.1604640

29. Gammoh, N.Z., and L. Rink. Zinc in Infection and Inflammation. Nutrients, 2017.9 (6):p.624. https://doi.org/10.3390/nu9060624

30. Hsu, R.H., et al., Genotypic and Phenotypic Correlations of Biotinidase Deficiency in the Chinese Population. Orphanet Journal of Rare Diseases, 2019. 14 (1): p.1–6. https://doi.org/10.1186/s13023-018-0992-2

31. Schwantje, M., et al., Genetic Defect of the Sodium-dependent Multivitamin Transporter: A Treatable Disease, Mimicking Biotinidase Deficiency. JIMD Reports , 2019. 48 (1): p.11–14. https://doi.org/10.1002/jmd2.12040.

32. Misir, R., R. Blair, and C.E. Doige, Development of a System for Clinical Evaluation of the Biotin Status of Sows. The Canadian Veterinary Journal = La Revue Veterinaire Canadienne, 1986. 27 (1):p. 6–12. PMID: 17422621

33. CDC. 1992. Recommendations for the use of folic acid to reduce the number of cases of spina bifida and other neural tube defects. MMWR Recomm Rep 41, 1–7.

34. SACN. 2017. Folic Acid: Updated Recommendations Issued by the Scientific Advisory Committee on Nutrition. London: Public Health England

35. FFI. 2019. Food Fortification Initiative. http://www.ffinetwork.org/global_progress/ (Accessed March 21, 2020).

36. Tinelli, C. et al., Hyperhomocysteinemia as a Risk Factor and Potential Nutraceutical Target for Certain Pathologies. Front Nutr, 2019. 6:p.49. https://doi.org/10.3389/fnut.2019.00049

37. Caffrey, A., et al., Maternal folate nutrition and offspring health: evidence and current controversies. Proc Nutr Soc, 2019. 78(2):p.208–220. https://doi.org/10.1017/S0029665118002689

38. Caffrey, A., et al., Gene specific DNA methylation in newborns in response to folic acid supplementation during the second and third trimesters of pregnancy: epigenetic analysis from a randomized controlled trial. Am J Clin Nutr, 2018. 107: 566–575. https://doi.org/10.1093/ajcn/nqx069

39. Braun, V.E. et al., Dietary Intakes of Folic Acid and Methionine in Early Childhood Are Associated with Body Composition at School Age. The Journal of Nutrition, 2015. 145(9): p. 2123–2129. https://doi.org/10.3945/jn.115.216283

40. Lykkesfeldt, J., and P. Tveden-Nyborg, The Pharmacokinetics of Vitamin C. Nutrients, 2019 11(10):p.2412. https://doi.org/10.3390/nu11102412

41. Ibrahim, B.S., et al., Beneficial effects of vitamin C treatment on pregnant rats exposed to formaldehyde: reversal of immunosuppression in the offspring. Toxicol Appl Pharmacol, 2016. 300:p.77–81. https://doi.org/10.1016/j. taap.2016.03.010

42. Wang, Y.Z., et al., Concentrations of antioxidant vitamins in maternal and cord serum and their effect on birth outcomes. J Nutr Sci Vitaminol (Tokyo), 2009. 55(1):p.1–8. https://doi.org/10.3177/jnsv.55.1

43. Ahn, Y., et al., Prenatal Vitamin C Status is Associated with Placental Apoptosis in Normal-term Human Pregnancies. Placenta, 2007. 28(1): p.31-38. https://doi.org/10.1016/j.placenta.2006.01.018

44. Saker, M., et al., Oxidant and antioxidant status in mothers and their newborns according to birthweight. Eur J Obstet Gynecol Reprod Biol, 2008. 141(2):p. 95–99. https://doi.org/10.1016/j.ejogrb.2008.07.013

45. Siega-Riz, A.M., et al., Vitamin C intake and the risk of preterm delivery. Am J Obstet Gynecol., 2003;189(2):p. 519–525. https://doi.org/10.1067/S0002-9378(03)00363-6

46. Hong, J., et al., Association of antioxidant vitamins and oxidative stress levels in pregnancy with infant growth during the first year of life. Public Health Nutrition, 2008; 11(10): p. 998-1005. https://doi.org/10.1017/S1368980007001322

47. Mercer, B.M., et al., The impact of vitamin C supplementation in pregnancy and in vitro upon fetal membrane strength and remodeling. Reprod Sci, 2010;17(7):p.685-95. https://doi.org/10.1177/1933719110368870

48. Jang, W., et al., Maternal fruit and vegetable or vitamin C consumption during pregnancy is associated with fetal growth and infant growth up to 6 months: results from the Korean Mothers and Children's Environmental Health (MOCEH) cohort study. Nutr J., 2018. 17(1):p105. https://doi.org/10.1186/s12937-018-0410-6

49. Hansen, S.T., et al., Maternal vitamin C deficiency does not reduce hippocampal volume and β-tubulin III intensity in prenatal Guinea pigs. Nutrition Research, 2016. 36(7):p.696-702. https://doi.org/10.1016/j.nutres.2016.03.004

50. Tveden-Nyborg, P., and J. Lykkesfeldt, Does vitamin C deficiency result in impaired brain development in infants? Redox Report, 2009. 14(1):p.2-6, https://doi.org/10.1179/135100009X392412

51. Tveden-Nyborg, P., et al. Maternal vitamin C deficiency during pregnancy persistently impairs hippocampal neurogenesis in offspring of Guinea pigs. PLoS One, 2012. 7:p.e48488. https://doi.org/10.1371/journal.pone.0048488

52. Ehrenstein, V., et al., Pregnancy-associated hypertensive disorders and adult cognitive function among Danish conscripts. Am. J. Epidemiol., 2009. 170(8):p. 1025–1031. https://doi.org/10.1093/aje/kwp223

53. Tuovinen, S., et al., Hypertensive disorders in pregnancy and intellectual abilities in the offspring in young adulthood: The Helsinki Birth Cohort Study. Ann. Med., 2012. 44:p.394–403. https://doi.org/10.3109/07853890.2011.573497

54. Many, A., et al. Neurodevelopmental and cognitive assessment of children born growth restricted to mothers with and without preeclampsia. Hypertens Pregnancy, 2003. 22(1):p. 25–29. https://doi.org/10.1081/PRG-120016791

55. Mikhail, M.S., et al., Preeclampsia and antioxidant nutrients—Decreased plasma levels of reduced ascorbic acid, alpha-tocopherol and beta-carotene in women with preeclampsia. Am. J. Obstet. Gynecol, 1994. 171:p. 150–157. https://doi.org/10.1016/0002-9378(94)90462-6

56. Takeshita, N., et al., Deficiency of ascorbic acid decreases the contents of tetrahydrobiopterin in the liver and the brain of ODS rats. Neurosci Lett, 2020. 715:p.134656. https://doi.org/10.1016/j.neulet.2019.134656

57. DiTroia, S.P., et al., Maternal vitamin C regulates reprogramming of DNA methylation and germline development. Nature, 2019. 573: p.271–275. https://doi.org/10.1038/s41586-019-1536-1

58. Cabezuelo, M.T., et al., Role of vitamin A in mammary gland development and lactation. Nutrients, 2020. 12(1):p.1–17. https://doi.org/10.3390/nu12010080

59. Basu, S., et al., Oral vitamin A supplementation in very low birth weight neonates: a randomized controlled trial. European Journal of Pediatrics, 2019. 178(8):p. 1255–1265. https://doi.org/10.1007/s00431-019-03412-w

60. Maia, S.B., et al., Vitamin a and pregnancy: A narrative review. Nutrients, 2019. 11(3); p.1–18. https://doi.org/10.3390/nu11030681

61. Dhruba, S., et al., Prevalence of vitamin D deficiency in pregnant women and their babies in Bhaktapur, Nepal. BMC Nutr, 2019; 5: p.31. https://doi.org/10.1186/s40795-019-0294-7

62. von Websky, K, et al., Impact of vitamin D on pregnancy-related disorders and on offspring outcome. J Steroid Biochem Mol Biol, 2018. 180:p.51–64. https://doi.org/10.1016/j.jsbmb.2017.11.008

63. Abbasian ,M., et al., Vitamin D Deficiency in Pregnant Women and Their Neonates. Glob J Health Sci, 2016. 8(9): p.83–90. https://doi.org/10.5539/gjhs.v8n9p83

64. Holick, M.F., et al., Evaluation, treatment, and prevention of vitamin D deficiency: an Endocrine Society clinical practice guideline. J Clin Endocrinol Metab, 2011. 96(7):p.1911-30. https://doi.org/10.1210/jcem.96.12.zeg3908

65. Marshall, I., R. Mehta, and A. Petrova, Vitamin D in the maternal-fetal-neonatal interface: clinical implications and requirements for supplementation. J Matern Fetal Neonatal Med, 2013. 26(7):633–638. https://doi.org/10.3109/147670 58.2012.746306

66. Aghajafari, F., et al., Association between maternal serum 25-hydroxyvitamin D level and pregnancy and neonatal outcomes: systematic review and meta-analysis of observational studies. BMJ, 2013. 346 :p.f1169. https://doi. org/10.1136/bmj.f1169

67. Cannell, J.J., Vitamin D and autism, what's new? Reviews in Endocrine and Metabolic Disorders, 2017. 18(2):p. 183–193. https://doi.org/10.1007/s11154-017-9409-0

68. Wagner, C.L., and B.W. Hollis, The Implications of Vitamin D Status During Pregnancy on Mother and her Developing Child. Front Endocrinol (Lausanne), 2018. 9:p.500. https://doi.org/10.3389/fendo.2018.00500

69. Ozdemir, A.A., and Y. Cag, Neonatal Vitamin D status and the risk of neonatal sepsis. Pak J Med Sci, 2019. 35(2):p. 420–425. https://doi.org/10.12669/pjms.35.2.342

70. Zhao, X., et al. Vitamin D Status among Young Children Aged 1-3 Years: A Cross-Sectional Study in Wuxi, China. PLoS One, 2015. 10(10): p. e0141595. https://doi.org/10.1371/journal.pone.0141595

71. Santamaria, C. et al., Prenatal vitamin D status and offspring's growth, adiposity and metabolic health: a systematic review and meta-analysis. British Journal of Nutrition, 2018. 119(03):p.310–319. https://doi.org/10.1017/ s0007114517003646

72. Luk, J., et al., Relevance of vitamin D in reproduction. Hum Reprod, 2012. 27(10):p. 3015-27. https://doi.org/10.1093/ humrep/des248

73. Dabrowski, F.A., B. Grzechocinska, and M. Wielgos, The role of vitamin D in reproductive health-- a Trojan Horse or the Golden Fleece? Nutrients, 2015. 7(6):p.4139-53. https://doi.org/10.3390/nu7064139

74. Merewood, A., et al., Association between vitamin D deficiency and primary cesarean section. J Clin Endocrinol Metab, 2009. 94(3): p. 940 –945. https://doi.org/10.1210/jc.2008-1217

75. Dawodu, A., and H. Akinbi, Vitamin D nutrition in pregnancy: current opinion. Int J Womens Health, 2013. 5:p.333–343. https://doi.org/10.2147/IJWH.S34032

76. Larqué, E., et al., Maternal and Foetal Health Implications of Vitamin D Status during Pregnancy. Annals of Nutrition and Metabolism, 2018. 72(3), p.179–192. https://doi.org/10.1159/000487370

77. Eremkina AK, Mokrysheva NG, Pigarova EA, Mirnaya SS. Vitamin D: effects on pregnancy, maternal, fetal and postnatal outcomes. Ter Arkh. 2018 Nov 22;90(10):115-127. https://doi.org/10.26442/terarkh20189010115-127

78. Shin, J.S., et al., Vitamin D effects on pregnancy and the placenta. Placenta, 2010. 31(12):p.1027-34. https://doi. org/10.1016/j.placenta.2010.08.015

79. Barrera, D., Vitamin D and Inflammatory Cytokines in Healthy and Preeclamptic Pregnancies. Nutrients, 2015. 7(8):p. 6465–6490. https://doi.org/10.3390/nu7085293

80. Grayson, R., and M. Hawison, Vitamin D and human pregnancy. Fetal and Maternal Medicine Review, 2011. 22(1):p.67–90. https://doi.org/10.1017/S0965539511000039

81. Behrouz, G.F., et al., Presence of auto-antibody against two placental proteins, annexin A1 and vitamin D binding protein, in sera of women with pre-eclampsia. J Reprod Immunol, 2013. 99(1-2):p.10-6. https://doi.org/10.1016/j. jri.2013.04.007

82. Viljakainen, H.T., et al., Maternal Vitamin D Status Determines Bone Variables in the Newborn, The Journal of Clinical Endocrinology & Metabolism, 2010. 95(4):p.1749–1757. https://doi.org/10.1210/jc.2009-1391

83. Cho, G.J., et al., Vitamin D deficiency in gestational diabetes mellitus and the role of the placenta. Am J Obstet Gynecol, 2013. 209(6):p.560.e1-8. https://doi.org/10.1016/j.ajog.2013.08.015

84. Wimalawansa, S.J., Associations of vitamin D with insulin resistance, obesity, type 2 diabetes, and metabolic syndrome. J Steroid Biochem Mol Biol, 2016. https://doi.org/10.1016/j.jsbmb.2016.09.017

85. Lu, M., et al., Association between vitamin D status and the risk of gestational diabetes mellitus: a meta-analysis. Arch Gynecol Obstet, 2016. 293(5):p.959-66. https://doi.org/10.1007/s00404-016-4010-4

86. Zhang, Q., et al., Effect of various doses of vitamin D supplementation on pregnant women with gestational diabetes mellitus: a randomized controlled trial. Exp Ther Med, 2016. 12:p.1889–1895. https://doi.org/10.3892/etm.2016.3515

87. Kim, I., et al., Association between vitamin D level at birth and respiratory morbidities in very-low-birth-weight infants. Korean J Pediatr, 2019. 62(5):p.166–172. https://doi.org/10.3345/kjp.2018.06632

88. Albertini, F., et al., Two cases of fractures in neonates associated with maternofetal vitamin D deficiency. Archives de Pédiatrie, 2019. 26(6):p.361-364. https://doi.org/10.1016/j.arcped.2019.06.004

89. Holick, M.F. Vitamin D Deficiency. N Engl J Med 2007; 357:p. 266-281. https://doi.org/10.1056/NEJMra070553

90. Cetinkaya, M., et al., Lower vitamin D levels are associated with increased risk of early-onset neonatal sepsis in term infants. J Perinatol, 2015. 35(1):39–45. https://doi.org/10.1038/jp.2014.146

91. Omran, A., et al., Maternal and neonatal vitamin D deficiency and transient tachypnea of the newborn in full term

neonates. J Perinat Med, 2018. 46(9):p.1057-1060. https://doi.org/10.1515/jpm-2017-0280

92. Saboute, M., et al., Investigation of association between maternal 25-OH vitamin D serum levels and neonatal early onset sepsis in newborns by evaluating key factors. Lipids Health Dis, 2019.18(1):p.153. https://doi.org/10.1186/s12944-019-1095-3

93. Çetinkaya, M., et al., Maternal/neonatal vitamin D deficiency: a risk factor for bronchopulmonary dysplasia in preterms?. J Perinatol, 2015. 35(10):p.813–817. https://doi.org/10.1038/jp.2015.88

94. Mohamed Hegazy, A., et al., Association between serum 25 (OH) vitamin D level at birth and respiratory morbidities among preterm neonates. J Matern Fetal Neonatal Med, 2018. 31(20): p.2649–2655. https://doi.org/10.1080/14767058 .2017.1350162

95. Zhao, X. et al., Maternal Vitamin D Status in the Late Second Trimester and the Risk of Severe Preeclampsia in Southeastern China. Nutrients, 2017. 9(2):p. 138. https://doi.org/10.3390/nu9020138

96. Regnault, T.R., et al., Placental development in normal and compromised pregnancies-- a review. Placenta, 2002. 23 Suppl A:S119-29. https://doi.org/10.1053/plac.2002.0792

97. Cosmi, E., et al., Consequences in infants that were intrauterine growth restricted. J Pregnancy, 2011. 2011:p.364381. https://doi.org/10.1155/2011/364381

98. Gezmish, O., and M.J. Black, Vitamin D Deficiency in Early Life and the Potential Programming of Cardiovascular Disease in Adulthood. Journal of Cardiovascular Translational Research, 2013. 6(4):p. 588–603. https://doi.org/10.1007/s12265-013-9475-y

99. Koster, M.P.H., et al., A compromised maternal vitamin D status is associated with congenital heart defects in offspring. Early Hum Dev, 2018. 117:p.50–56. https://doi.org/10.1016/j.earlhumdev.2017.12.011

100. Erkkola M., M. Kaila, B.I. Nwaru, C. Kronberg-Kippilä, S. Ahonen, J. Nevalainen, R. Veijola, J. Pekkanen, J. Ilonen, O. Simell, M. Knip, S.M. Virtanen, Maternal vitamin D intake during pregnancy is inversely associated with asthma and allergic rhinitis in 5-year-old children, Clin. Exp. Allergy 39 (2009) 875–882, http://dx.doi.org/10.1111/j.1365-2222.2009.03234.x

101. Wjst, M., Is vitamin D supplementation responsible for the allergy pandemic? Curr. Opin. Allergy Clin. Immunol, 2012. 12(3): p. 257–262. https://doi.org/10.1097/ACI.0b013e3283535833

102. Weisse K., et al., Maternal and newborn vitamin D status and its impact on food allergy development in the German LINA cohort study. Allergy, 2013. 68(2):p. 220–28 https://doi.org/10.1111/all.12081

103. Patrick, R.P., and B.N. Ames, Vitamin D Hormone regulates serotonin synthesis. Part 1: relevance for autism. FASEB J, 2014. 28(6): p.2398–2413. https://doi.org/10.1096/fj.13-246546

104. Huang, Y.N., et al., 1,25- dihydroxyvitamin D3 attenuates endotoxin-induced production of inflammatory mediators by inhibiting MAPK activation in primary cortical neuron-glia cultures. J Neuroinflammation, 2015. 12:p.147. https://doi.org/10.1186/s12974-015-0370-0

105. Arnson, Y., et al., Vitamin D inflammatory cytokines and coronary events: a comprehensive review. Clin Rev Allergy Immunol, 2013; 45(2):p. 236–47. https://doi.org/10.1007/s12016-013-8356-0

106. Krakowiak, P., et al., Neonatal Cytokine Profiles Associated With Autism Spectrum Disorder. Biol Psychiatry, 2017. 81(5):p.442-451. https://doi.org/10.1016/j.biopsych.2015.08.007

107. Maladkar, M., S. Sankar, and K. Kamat, Vitamin D efficiency in pregnancy: An updated viewpoint in Indian scenario. Int. J. Clinical Medicine, 2015. 6(3):p.204-16. https://org.doi/10.4236/ijcm.2015.63026

108. Stubbs, G., K. Henley, J. Green, Autism: Will vitamin D supplementation during pregnancy and early childhood

reduce the recurrence rate of autism in newborn siblings?. Med Hypotheses, 2016. 88:74–78. https://doi.org/10.1016/j. mehy.2016.01.015

109. Mossin, M.H., etal., Inverse associations between cord vitamin D and attention deficit hyperactivity disorder symptoms: A child cohort study. Aust N Z J Psychiatry, 2017. 51(7):p.703–710. https://doi.org/10.1177/0004867416670013

110. Sahin, N., et al., Vitamin D and vitamin D receptor levels in children with attention-deficit/hyperactivity disorder. Neuropsychiatr Dis Treat, 2018. 14:p.581–585. https://doi.org/10.2147/NDT.S158228

111. Khoshbakht, Y., R. Bidaki, and A. Salehi-Abargouei, Vitamin D Status and Attention Deficit Hyperactivity Disorder: A Systematic Review and Meta-Analysis of Observational Studies. Adv Nutr, 2018. 9(1):p.9–20. https://doi.org/10.1093/advances/nmx002

112. Kazemi, A., et al., High prevalence of vitamin D deficiency among pregnant women and their newborns in an Iranian population. J Womens Health (Larchmt), 2009. 18(6):835–839. https://doi.org/10.1089/jwh.2008.0954

113. Elshorbagy, H.H., et al., Impact of Vitamin D Supplementation on Attention-Deficit Hyperactivity Disorder in Children. Ann Pharmacother., 2018; 52(7):p.623–631. https://doi.org/10.1177/1060028018759471

114. Kinney, D.K., et al., Relation of schizophrenia prevalence to latitude, climate, fish consumption, infant mortality, and skin color: a role for prenatal vitamin d deficiency and infections?. Schizophr Bull., 2009. 35(3):p. 582–595. https://doi.org/10.1093/schbul/sbp023

115. Kabataş, E.U., et al., Relationship between serum 25-hydroxy vitamin D levels and retinopathy of prematurity. Scottish Medical Journal, 2017. 62(4):p. 129–135. https://doi.org/10.1177/0036933017701867

116. Helou, M., et al., Vitamin D Deficiency in Children With Cancer. Journal of Pediatric Hematology/Oncology, 2014. 36(3), p. 212–217. https://doi.org/10.1097/mph.0b013e31829f3754

117. Brion, L.P., E.F. Bell, and T.S. Raghuveer, Vitamin E supplementation for prevention of morbidity and mortality in preterm infants. Cochrane Database of Systematic Reviews, 2003. Issue 4. Art. No.: CD003665. https://doi. org/10.1002/14651858.CD003665

118. Ben Hamida, M., et al., Friedreich's ataxia phenotype not linked to chromosome 9 and associated with selective autosomal recessive vitamin E deficiency in two inbred Tunisian families. Neurology, 1993; 43(11):p.2179-83. https://doi.org/10.1212/wnl.43.11.2179

119. Schuelke, M., et al., Treatment of ataxia in isolated vitamin E deficiency caused by alpha-tocopherol transfer protein deficiency. J Pediatr 1999; 134:p.240-44. https://doi.org/10.1016/S0022-3476(99)70424-5

120. El Euch-Fayache, G., et al., Molecular, clinical and peripheral neuropathy study of Tunisian patients with ataxia with vitamin E deficiency. Brain, 2014. 137:p.402. https://doi.org/10.1093/brain/awt339

121. Schuelke, M. Ataxia with Vitamin E Deficiency. 2005 May 20 [Updated 2013 Jun 27]. In: Pagon RA, Adam MP, Ardinger HH, et al., editors. GeneReviews® [Internet]. Seattle (WA): University of Washington, Seattle; 1993-2014. Available from: http://www.ncbi.nlm.nih.gov/books/NBK1241/

122. van de Warrenburg, B.P., et al., EFNS/ENS Consensus on the diagnosis and management of chronic ataxias in adulthood. Eur J Neurol, 2014. 21(4):p.552-562. https://doi.org/10.1111/ene.12341

123. Oski, F.A., and L. Barness, Vitamin E deficiency: a previously unrecognized cause of hemolytic anemia in the premature infant. J Pediatr ,1967; 70:p.211.

124. Natta, C., and L. Machlin, Plasma levels of tocopherol in sickle cell anemia subjects. Am J Clin Nutr, 1979. 32:p.1359. https://doi.org/10.1093/ajcn/32.7.1359

125. Ray, D., et al., Antioxidant vitamin levels in sickle cell disorders. Natl Med J India, 2007; 20:p.11. PMID:

17557515

126. Walter, P.B., et al., Oxidative stress and inflammation in iron-overloaded patients with beta-thalassaemia or sickle cell disease. Br J Haematol., 2006. 135(2):p. 254-263. https://doi.org/10.1111/j.1365-2141.2006.06277.x

127. Mihatsch, W.A., et al., ESPGHAN Committee on Nutrition. Prevention of Vitamin K Deficiency Bleeding in Newborn Infants: A Position Paper by the ESPGHAN Committee on Nutrition. J. Pediatr. Gastroenterol. Nutr., 2016. 63(1):p.123-9. https://doi.org/10.1097/MPG.00000000001232

128. Fusaro, M., et al., Vitamin K plasma levels determination in human health. Clin. Chem. Lab. Med., 2017. 55(6):p. 789-799. https://doi.org/10.1515/cclm-2016-0783

129. Wen, L., et al., Vitamin K dependent proteins involved in bone and cardiovascular health (Review). Mol Med Rep, 2018. 18(1):p.3-15. https://doi.org/10.3892/mmr.2018.8940

130. Riphagen, I.J., et al., Prevalence and Effects of Functional Vitamin K Insufficiency: The PREVEND Study. Nutrients, 2017. 9(12). https://doi.org/10.3390/nu9121334

131. Napolitano, M., G. Mariani, and M. Lapecorella, Hereditary combined deficiency of the vitamin K-dependent clotting factors. Orphanet J Rare Dis, 2010.5:p.21. https://doi.org/10.1186/1750-1172-5-21

132. Marchili, M.R., et al., Vitamin K deficiency: a case report and review of current guidelines. Ital J Pediatr, 201. 44(1):p.36. https://doi.org/10.1186/s13052-018-0474-0

133. Araki S, and A. Shirahata, Vitamin K Deficiency Bleeding in Infancy, Nutrients, 2020. 12(3):p.780. https://doi. org/10.3390/nu12030780

134. Shearer, M.J, Vitamin K deficiency bleeding (VKDB) in early infancy. Blood Rev, 2009.23(2):p.49-59. https://doi. org/10.1016/j.blre.2008.06.001

135. Mousa, A., A. Naqash, and S. Lim. Macronutrient and Micronutrient Intake during Pregnancy: An Overview of Recent Evidence. Nutrients, 2019. 11 (2): p.443. https://doi.org/10.3390/nu11020443

136. Olausson, H., et al., Calcium Economy in Human Pregnancy and Lactation. Nutrition Research Reviews, 2012. 25 (1): p.40–67. https://doi.org/10.1017/S0954422411000187

137. Almaghamsi, A., M.H. Almalki, and B.M. Buhary. Hypocalcemia in Pregnancy: A Clinical Review Update. Oman Medical Journal, 2018. 33 (6): p.453–62. https://doi.org/10.5001/omj.2018.85

138. Palmsten, K., S.L. Buka, and K.B. Michels. Maternal Pregnancy-Related Hypertension and Risk for Hypertension in Offspring Later in Life. Obstetrics and Gynecology, 2010. 116 (4): 858–64. https://doi.org/10.1097/AOG.0b013e3181f3a1f9

139. Djagbletey, R., et al., Serum Calcium and Magnesium Levels in Normal Ghanaian Pregnant Women: A Comparative Cross-Sectional Study. Open Access Macedonian Journal of Medical Sciences, 2018. 21;6(11):p.2006-2011. https://pubmed.ncbi.nlm.nih.gov/30559851/

140. Takaya, J., Small for Gestational Age and Magnesium: Intrauterine Magnesium Deficiency May Induce Metabolic Syndrome in Later Life. AIMS Public Health, 2015. 2 (4): p.793–803. https://doi.org/10.3934/publichealth.2015.4.793

141. Maggini, S., A. Pierre, and P.C. Calder, Immune Function and Micronutrient Requirements Change over the Life Course. Nutrients, 2018. 10 (10): p.1531. https://doi.org/10.3390/nu10101531

142. Das, S., J. et al., Dietary Magnesium, Vitamin D, and Animal Protein Intake and Their Association to the Linear Growth Trajectory of Children from Birth to 24 Months of Age: Results From MAL-ED Birth Cohort Study Conducted in Dhaka, Bangladesh. Food and Nutrition Bulletin, 2020. 3795721198: 1–11. https://doi.org/10.1177/0379572119892408

143. Caulfield, L.E., et al., Potential Contribution of Maternal Zinc Supplementation during Pregnancy to Maternal and

Child Survival. The American Journal of Clinical Nutrition, 1998. 68 (Suppl 2): 499S-508S. https://doi.org/10.1093/ ajcn/68.2.499S

144. Ota, E., et al., Zinc Supplementation for Improving Pregnancy and Infant Outcome. Cochrane Database of Systematic Reviews, 2012. CD000230 (7). https://doi.org/10.1002/14651858.CD000230.pub5

145. Gebreselassie, S.G., and F.E. Gashe. A Systematic Review of Effect of Prenatal Zinc Supplementation on Birthweight: Meta-Analysis of 17 Randomized Controlled Trials. Journal of Health, Population and Nutrition (JHPN), 2011. 29 (2): p.134–40. https://doi.org/10.3329/JHPN.V29I2.24

146. Chaffee, B.W., and J.C. King, Effect of Zinc Supplementation on Pregnancy and Infant Outcomes: A Systematic Review. Paediatric and Perinatal Epidemiology, 2012. 26 (Suppl. 1): p.118–37. https://doi.org/10.1111/j.1365-3016.2012.01289.x

147. Chowanadisai, W., S.L. Kelleher, and B. Lönnerdal, Maternal Zinc Deficiency Reduces NMDA Receptor Expression in Neonatal Rat Brain, Which Persists into Early Adulthood. Journal of Neurochemistry, 2005. 94 (2): p.510–19. https://doi.org/10.1111/j.1471-4159.2005.03246.x

148. Yu, X., et al., Effects of Maternal Mild Zinc Deficiency and Different Ways of Zinc Supplementation for Offspring on Learning and Memory. Food & Nutrition Research, 2016. 60 (1): p.29467. https://doi.org/10.3402/fnr.v60.29467

149. Sauer, A.K., and A.M. Grabrucker, Zinc Deficiency During Pregnancy Leads to Altered Microbiome and Elevated Inflammatory Markers in Mice. Frontiers in Neuroscience, 2019. 13: p.1295. https://doi.org/10.3389/fnins.2019.01295

150. Jarosz, M., et al., Antioxidant and Anti-Inflammatory Effects of Zinc. Zinc-Dependent NF-KB Signaling. Inflammopharmacology, 2017. 25: p.11–24. https://doi.org/10.1007/s10787-017-0309-4

151. Ackland, M.L., and A.A. Michalczyk, Zinc and Infant Nutrition." Archives of Biochemistry and Biophysics, 2016. 611: p.51–57. https://doi.org/10.1016/j.abb.2016.06.011

152. Brown, K.H., et al., Preventive Zinc Supplementation among Infants, Preschoolers, and Older Prepubertal Children. Food and Nutrition Bulletin, 2009. 30 (Suppl.1). https://doi.org/10.1177/15648265090301s103

153. Qazi, I.H., et al., Selenium, Selenoproteins, and Female Reproduction: A Review. Molecules, 2018. 23 (12): E3053. https://doi.org/10.3390/molecules23123053

154. Mesdaghinia, E., Azam Rahavi, Fereshteh Bahmani, Nasrin Sharifi, and Zatollah Asemi. 2017. "Clinical and Metabolic Response to Selenium Supplementation in Pregnant Women at Risk for Intrauterine Growth Restriction: Randomized, Double-Blind, Placebo-Controlled Trial." Biological Trace Element Research 178 (1): 14–21. https://doi. org/10.1007/s12011-016-0911-0

155. Skröder, H.M., et al., Selenium Status in Pregnancy Influences Children's Cognitive Function at 1.5 Years of Age. Clinical Nutrition, 2015. 34 (5): p. 923–30. https://doi.org/10.1016/j.clnu.2014.09.020

156. Varsi, K., et al., Impact of Maternal Selenium Status on Infant Outcome during the First 6 Months of Life. Nutrients, 2017. 9 (5): E487. https://doi.org/10.3390/nu9050486

157. Skrökler, H., et al., Early-Life Selenium Status and Cognitive Function at 5 and 10 Years of Age in Bangladeshi Children. Environmental Health Perspectives, 2017. 125 (11): p.117003. https://doi.org/10.1289/EHP1691

158. Muntau, A.C., et al., Age-Related Reference Values for Serum Selenium Concentrations in Infants and Children. Clinical Chemistry, 2002. 48 (3): p.555–60. https://doi.org/10.1093/clinchem/48.3.555

159. Dror, Daphna K, and Lindsay H Allen. 2018. "Overview of Nutrients in Human Milk." Advances in Nutrition (Bethesda, Md.) 9 (suppl_1): 278S-294S. https://doi.org/10.1093/advances/nmy022.

160. Daniels L, Gibson RA, Simmer K, Van Dael P, Makrides M. Selenium status of term infants fed selenium-

supplemented formula in a randomized dose-response trial. Am J Clin Nutr. 2008;88(1):70-76. https://doi.org/10.1093/ ajcn/88.1.70

161. Brannon, P. M., and C.L. Taylor, Iron Supplementation during Pregnancy and Infancy: Uncertainties and Implications for Research and Policy. Nutrients, 9 (12): p.1327. https://doi.org/10.3390/nu9121327

162. Parisi, F., et al., Micronutrient Supplementation in Pregnancy: Who, What and How Much?. Obstetric Medicine, 2019. 12 (1): p.5–13. https://doi.org/10.1177/1753495X18769213

163. Domellöf, M., et al., Iron Requirements of Infants and Toddlers. Journal of Pediatric Gastroenterology and Nutrition, 2014. 58 (1): p.119–29. https://doi.org/10.1097/MPG.000000000000206

164. Hajianfar, H., et al., The Association between Maternal Dietary Iron Intake during the First Trimester of Pregnancy with Pregnancy Outcomes and Pregnancy-Related Complications. Clinical Nutrition Research, 2020. 9 (1): p.52–62. https://doi.org/10.7762/cnr.2020.9.1.52

165. Milman, N., et al., Supplementation during Pregnancy: Beliefs and Science. Gynecological Endocrinology, 2016.32 (7): p.509–16. https://doi.org/10.3109/09513590.2016.1149161

166. Juul, S.E., R.J. Derman, and M. Auerbach. Perinatal Iron Deficiency: Implications for Mothers and Infants. Neonatology, 2019. 115 (3): p.269–74. https://doi.org/10.1159/000495978

167. Georgieff, M.K., Long-Term Brain and Behavioral Consequences of Early Iron Deficiency. Nutrition Reviews, 2011. 69 (Suppl. 1): S43–48. https://doi.org/10.1111/j.1753-4887.2011.00432.x

168. Alwan, N., and H.Hamamy, Maternal Iron Status in Pregnancy and Long-Term Health Outcomes in the Offspring. Journal of Pediatric Genetics, 2015. 4 (2): p.111–23. https://doi.org/10.1055/s-0035-1556742

169. Martin CR, Ling PR, Blackburn GL. Review of Infant Feeding: Key Features of Breast Milk and Infant Formula. Nutrients. 2016;8(5):279. Published 2016 May 11. doi:10.3390/nu8050279

170. Innis, S.M., Dietary (n-3) fatty acids and brain development. The Journal of nutrition, 2007. 137(4):p.855-9. https://doi.org/10.1093/jn/137.4.855

171. Al M.D., A.C. van Houwelingen, and G. Hornstra, Long-chain polyunsaturated fatty acids, pregnancy, and pregnancy outcome. The American journal of clinical nutrition. Am J Clin Nutr, 2000 Jan;71(1):p.285S-91S. https://doi. org/10.1093/ajcn/71.1.285s

172. Cetin, I., G. Alvino, and M. Cardellicchio. Long Chain Fatty Acids and Dietary Fats in Fetal Nutrition. In Journal of Physiology, 2009. 587:p.3441–51. https://doi.org/10.1113/jphysiol.2009.173062

173. Rosolem S., et al., Optimization during the First Thousand Days of Child through Dietary Supplement with Lc-Pufas: Systematic Review of the Literature. Maternal and Pediatric Nutrition, 2017. 03 :p.01. https://doi.org/10.4172/2472-1182.1000120

174. Haggarty P. Fatty Acid Supply to the Human Fetus." Annual Review of Nutrition 30 (1): 237–55. https://doi. org/10.1146/annurev.nutr.012809.104742

175. Herrera E, H. Ortega-Senovilla, Maternal Lipid Metabolism during Normal Pregnancy and Its Implications to Fetal Development. Clinical Lipidology, 2010. 5:6: p.899-911. https://doi.org/10.2217/clp.10.64

176. Hsueh ,T.Y., J.I. Baum, and Y. Huang, Effect of Eicosapentaenoic Acid and Docosahexaenoic Acid on Myogenesis and Mitochondrial Biosynthesis during Murine Skeletal Muscle Cell Differentiation. Front Nutr, 2018; p.5:15. https://doi.org/10.3389/fnut.2018.00015

177. Quin, C., et al., Omega-3 Polyunsaturated Fatty Acid Supplementation during the Pre and Post-Natal Period: A Meta-Analysis and Systematic Review of Randomized and Semi-Randomized Controlled Trials. Journal of Nutrition

and Intermediary Metabolism, 2016. 5:p.34-54. https://doi.org/10.1016/j.jnim.2016.04.005.

178. Moon, K., et al., Longchain Polyunsaturated Fatty Acid Supplementation in Preterm Infants. Cochrane Database of Systematic Reviews, 2016. 12: p.8–26. https://doi.org/10.1002/14651858.CD000375.pub5.

179. Shulkin, M., et al., N-3 Fatty Acid Supplementation in Mothers, Preterm Infants, and Term Infants and Childhood Psychomotor and Visual Development: A Systematic Review and Meta-Analysis. Journal of Nutrition, 2018. 148 (3): p.409–18. https://doi.org/10.1093/jn/nxx031

180. Wang, Q, Q. Cui, and C.Yan, The Effect of Supplementation of Long-Chain Polyunsaturated Fatty Acids during Lactation on Neurodevelopmental Outcomes of Preterm Infant from Infancy to School Age: A Systematic Review and Meta-Analysis. Pediatric Neurology, 2016: 59: p.54-61.e1. https://doi.org/10.1016/j.pediatrneurol.2016.02.017

181. Lapillonne, A., and S. J. Moltu, Long-Chain Polyunsaturated Fatty Acids and Clinical Outcomes of Preterm Infants. Annals of Nutrition and Metabolism, 2016. 69 (1): p.36–44. https://doi.org/10.1159/000448265.

182. Chatzi, L., et al., Mediterranean Diet in Pregnancy Is Protective for Wheeze and Atopy in Childhood. Thorax, 2008; 63 (6): p.507–13. https://doi.org/10.1136/thx.2007.081745

183. Furuhjelm, C., et al., Fish Oil Supplementation in Pregnancy and Lactation May Decrease the Risk of Infant Allergy. Acta Paediatrica, International Journal of Paediatrics. 2009; 98 (9): p.1461–67. https://doi.org/10.1111/j.1651-2227.2009.01355.x.

184. Hirata, S., et al., Maternal Ω 3 Docosapentaenoic Acid Inhibits Infant Allergic Dermatitis through TRAIL-expressing Plasmacytoid Dendritic Cells in Mice, 2020.75:p.1939–1955. https://doi.org/10.1111/all.14217.

185. Mun, J.G., et al., Choline and DHA in Maternal and Infant Nutrition: Synergistic Implications in Brain and Eye Health. Nutrients, 2019. 11 (5):p.1125. https://doi.org/10.3390/nu11051125.

186. European Food Safety Authority (EFSA). Scientific Opinion on Dietary Reference Values for Fats, Including Saturated Fatty Acids, Polyunsaturated Fatty Acids, Monounsaturated Fatty Acids, Trans Fatty Acids, and Cholesterol. EFSA Journal, 2016. 8 (3): p.1–107. https://doi.org/10.2903/j.efsa.2010.1461

187. Speer, M.E., Health Literacy and Child Health Outcomes:From Prenatal to Birth and Infant Stages. In: Connelly RA, Speer ME, editors. Health Literacy and Child Health Outcomes: Promoting Effective Health Communication Strategies to Improve Quality of Care. Sp

188. Kaya Senol, D., I. Gol, and S.A. Ozkan, The Effect of Health Literacy Levels of Pregnant Women on Receiving Prenatal Care: A Cross-Sectional Descriptive Study. Int J Caring Sci, 2019. 12 (3):p.1717

189. Wilhelmova, R., D. Hruba, and L. Vesela, Key determinants influencing the health literacy of pregnant women in the Czech Republic. Zdr Varst, 2015. 54(1):27–36. https://doi.org/10.1515/sjph-2015-0004

190. Charoghchian Khorasani, E., N. Peyman, and H. Esmaily, Relations between Breastfeeding Self-efficacy and Maternal Health Literacy among Pregnant Women. 2017 Journal of Evidence-Based Care. 2017;6(4):p.18-25. https://doi.org/10.22038/ebcj.2016.7986

191. Koletzko, B., et al., Nutrition During Pregnancy, Lactation and Early Childhood and its Implications for Maternal and Long-Term Child Health: The Early Nutrition Project Recommendations. Ann Nutr Metab. 2019;74(2):p.93-106. https://doi.org/10.1159/000496471

192. Imdad, A., et al., Prenatal Nutrition and Nutrition in Pregnancy: Effects on Long-Term Growth and Development. In: Early Nutrition and Long-Term Health: Mechanisms, Consequences, and Opportunities. Elsevier Inc.; 2017. p. 3–24. https://doi.org/10.1016/B978-0-08-100168-4.00001-X

193. Ho, A., A.C. Flynn, and D. Pasupathy, Nutrition in pregnancy. Obstetrics, Gynaecology and Reproductive Medicine. Churchill Livingstone, 2016. 26(9):259–264. https://doi.org/10.1016/j.ogrm.2016.06.005

194. Sørensen, K., et al., Health literacy and public health: A systematic review and integration of definitions and models. BMC Public Health, 2012. 12: p.1–13. https://doi.org/10.1186/1471-2458-12-80

195. Velardo, S., The Nuances of Health Literacy, Nutrition Literacy, and Food Literacy. J Nutr Educ Behav, 2015. 47(4):p.385-389.e1. https://doi.org/10.1016/j.jneb.2015.04.328

196. van der Heide, I., et al., The Relationship Between Health, Education, and Health Literacy: Results From the Dutch Adult Literacy and Life Skills Survey. J Health Commun, 2013.18(sup1):p.172–84. https://doi.org/10.1080/10810730. 2013.825668

197. WHO. WHO recommendations on antenatal care for a positive pregnancy experience. Luxembourg; 2016.

198. WHO. Nutrition counselling during pregnancy [Internet]. [cited 2020 Mar 27].